Cytoarchitecture of cortex imaged by **Optical Coherence Tomography**

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Introduction

Brodmann areas (BA) differ from one another in cytoarchitecture [3]. While brain mapping has improved with ex vivo MRI and histological validation, the resolution of MRI has limits that constrain our ability to visualize these features, even ex vivo. Moreover, refined localization of brain areas is critical for brain areas is critical for brain areas such as Alzheimer's disease. Although in vivo biomarkers exist, many definitive diagnoses still rest on autopsy evaluation of histological tissue. Histology is labor intensive and staining, which render registration across slices and to other modalities difficult. We propose to use Optical Coherence Tomography (OCT) to image the cortical architecture in the medial temporal lobe (MTL), at the entorhinal and perirhinal boundary (BA 28 and 35 respectively) and in the temporal isocortex. The acquisition is performed directly on the blockface to overcome registration issues. Perirhinal and their boundaries have been delineated in ex vivo MRI model [4, 1]. Here, we build on this model to distinguish isocortical boundaries that are typically more subtle than mesocortical boundaries, and not easily visible with MRI.

Methods and Materials

Optical coherence tomography

OCT is an optical technique providing **3D high resolution** images up to several hundreds of microns in depth in biological tissue [5]. This technique avoids cutting, mounting or staining and greatly reduces deformations in the tissue. OCT detects differences in refractive index between tissues types, e.g. cell bodies and myelinated fibers.

Temporal Isocortex

The second experiment involves 5 isocortical samples (mean age 65.5, 2F/2M, 1 unknown). The most distiguishable architectonic features were manualy drawn on both Nissl and OCT images: the pial surface (PS), the gray/white matter boundary (GWB) and cortical layers (CL). Fig. 3A and 3B show the manual labeling on the Nissl and OCT images, respectively.







Tissue Preparation

Temporal lobe samples were used for this study: two sample of MTL containing the entorhinal and perirhinal cortex boundary (BA 28 and 35 respectively) and five samples of isocortex (BA 36, 20, 21 and 22). The samples were fixed and sectioned to collect 50μ m thick slices for histological staining. The blockface was photographed after each section. The slices were then hand mounted and stained with thionin. The remaining flat-faced tissue block was imaged with OCT.

Qualitative comparison

Fig. 1 shows the blockface (A), Nissl stain (B) and OCT (C) images of one isocortical sample. Nissl and OCT have inversed contrast. The Nissl shows distortions due to histological protocol and tissue integrity, e.g. tissue rips, tissue overlaps or wide gaps between gyri (Fig. 1B, arrows). Distortions can impair registration to blockface image or to MRI data. OCT shows minimal if any distortions.



Fig. 3: Manual labeling of the cortical landmarks on co-registered Nissl stain (A) and OCT images (B). The drawn lines are the PS (magenta), the GWB (red) and the differents CL: layer II (dark blue), layer III (cyan), layer IV (green) and layer V (yellow). C: Thickness from GWB to CL.

The plot (Fig. 3C) shows the thickness between GWB and CL of the sample presented on Fig. 3A and 3B. The thicknesses are symbolized by + for the Nissl stains and by \circ for the OCT. Qualitatively, the thicknesses are in good accordance. The Pearson's correlation ρ between the corresponding lines for both modalities was calculated for each sample, as well as their p-value. The mean Pearson's correlation was $ho=0.84\pm0.16$ with a very small p-value.

We have demonstrated that OCT labeling is as reliable as Nissl labeling. The labels drawn by two independent observers for each modality show no significant differences, evaluated by the Haussdorf distance and the median distance for each pair of lines (Fig. 4).



Fig. 1: A: Blockface, B: Nissl stain and C: OCT images of one of the isocortex samples.

Entorhinal and Perirhinal cortex boundary [2]

Fig. 2A and 2B show the OCT images (AIP and maximum intensity projection MIP, respectively) where the boundary between entorhinal (EC) and perirhinal (PC) cortex is observed as well as perirhinal subdivisions, areas 35a and 35b. In OCT (Fig. 2A), neuron dense areas appear white, in contrast to the Nissl stained section (Fig. 2C). Neurons are particularly notable in layers II and IV in EC and layers II and III in PC (individual neurons visible on the MIP in Fig. 2B). The oblique layer routinely observed in area 35 showed dark signal.





Fig. 4: Statistical analysis for the inter-observer reliability. A: the Haussdorff distance, B: the median minimal distance. Wilcoxon signed rank test is shown above the bracket by *, if p <= 0.05.

Advantages of OCT

Independent of tissue

The histological process and the ex vivo MRI are influenced by the tissue integrity and handling (aging, fixation, post mortem interval...) and/or affinity to dyes

Registration to blockface and MRI



deformations due to hand mounting or due to dehydration). Due to this, registration to the blockface images can be compromised (Fig. 5 left, arrow). Alternatively, OCT allows to image directly on the tissue block so that only minor distortions occur (Fig. 5, right).

Histology protocol suffers from distortions (rips,

Fig. 5: Registration to **blockface** with **Nissl** (left) and **OCT** (right).

► Cyto- and myelo- architectonic structures in cortical ribbon, as well as fibers in the white matter¹



Fig. 2: A: AIP, B: MIP and C: Nissl stain of MTL sample at the border of EC and PC.

Fig. 6: Overlay of MIP and AIP. A: Cyto- and myelo- architecture in the gray matter and B: connectivity in the white matter.

► 3D reconstruction

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Conclusions and Future Works

OCT is a promising tool for the localization of cortical boundaries. Indeed, in OCT data, especially the AIP, the laminar structure is similar to the one obtained by Nissl stain. However, contrary to histological staining, the tissue block is imaged, reducing considerably the deformations. Here 2D images were used to validate the technique but a future application will provide a 3D reconstruction of a volume of several cm³ by adding a vibrotome to our setup [6]. The OCT cortical segmentation could be used as a training set for *in vivo* MRI boundary detection.

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